

Fraunhofer
IME

Udo Hommen¹, Kerstin Derz¹, Elke Eilebrecht¹, Kirsten Germing¹, Andreas Schiermeyer², Sebastian Eilebrecht¹
¹ Fraunhofer IME, Schmallenberg, Germany, ² Fraunhofer IME, Aachen, Germany

- RNA interference (RNAi) means gene silencing in organisms caused by specific double-stranded RNA (dsRNA) or other small RNA molecules
- RNAi is a promising approach for crop protection, due to expected low environmental persistence of RNA molecules and potentially high specificity
- While genetically modified plants using RNAi against insect pests have not applied for cultivation in the EU, first RNAi plant protection products are expected to seek approval in the EU in the near future

- Summarize methods to analyse environmental fate and hazards of small RNA molecules used for RNAi
- Compare ERA frameworks for GMP and PPP
- Identify knowledge gaps

- Search in WebofScience and Pubmed plus inverse snowballing
- Many reviews but only limited comprehensive data sets on fate, exposure and hazard
- Most detailed documents:
 - Christiaens *et al.* (2018): Focus on RNAi GMP
 - OECD (2020): Focus on RNAi PPP
- Only two environmental risk assessments for insecticidal RNAi (conducted for approval in the USA)
 - MON 87411 (RNAi GMP, stacked maize)
 - *Calantha* (RNAi Spray)
- Acetamiprid 20 SG as chemical pesticide (neonicotinoid) for comparison

- *'Pathway to harm'* (adapted from Roberts et al. 2015)
 1. Introduction into environment
 2. Uptake by non-target organism
 3. No degradation after consumption
 4. Activation of RNAi machinery
 5. Sequence dependent (gene silencing) or sequence independent effect
 6. Effects on viability of the organism (surv, dev, repro)
 7. Effects relevant for Specific Protection Goals

```
graph LR; 1[1. Introduction into environment] --> 2[2. Uptake by non-target organism]; 2 --> 3[3. No degradation after consumption]; 3 --> 4[4. Activation of RNAi machinery]; 4 --> 5[5. Sequence dependent (gene silencing) or sequence independent effect]; 5 --> 6[6. Effects on viability of the organism (surv, dev, repro)]; 6 --> 7[7. Effects relevant for Specific Protection Goals];
```

	 RNAi GMP, e.g. MON87411	 RNAi spray, e.g. Calantha	 Chemical insecticide, e.g. Acetamiprid 20 SG
 Fate	dsRNA produced by crop Conc. in tissues? Long-term availability Low off-field exposure (debris, pollen)	Short-term availability on surfaces Uptake by crop? Relevance of drift, run-off and drainage to soil, off-field areas and surface waters	Substance and formulation specific fate and exposure
 Exposure	 Almost only via diet	 Via contact (overspray, surfaces, soil, water) and diet	
 Mode of action	 Gene silencing => only one target pest, high specificity Sequence independent effects less relevant	 E.g. neurotoxic often several target pests low specificity	
 Hazard	 Bioinformatics to identify low and high sensitive taxa Focussed bioassays Species selection, protocol modifications Field tests	 Standard bioassays / data requirements Higher tier tests	
 Risk	Case specific, e.g. levels of concern	Uniform principles using risk quotients or similar approaches	

- RNAi approaches can provide alternatives to chemical PPP due to often lower persistence of the active substances (short dsRNA molecules) and higher specificity
- RNAi PPP most likely to enter EU market
- EU regulation and frameworks for chemical PPP also applicable to RNAi PPP but some adaptations are needed
- Bioinformatics supports selection of test species but bioassays are still needed

- Consider 'pseudo-persistence' of RNA produced by GMP
- Develop refined exposure models, considering e.g. release of dsRNA from crop and food chain transfer
- Consider PPP formulation effects on fate of RNA
- Follow the 'need to know' approach of the EU commission in problem formulation for low risk PPP (e.g. check need of vertebrate testing)
- Adapt test protocols (dietary exposure, prolonged duration, sublethal effects, additional test species)

Christiaens et al. (2020). Double-Stranded RNA Technology to Control Insect Pests: Current Status and Challenges. *Frontiers in Plant Science*, 11, 451.
<https://doi.org/10.3389/fpls.2020.00451>

OECD (2020). Considerations for the Environmental Risk Assessment of the Application of Sprayed or Externally Applied ds-RNA-Based Pesticides. Series on Pesticides and Biocides. OECD. <https://doi.org/10.1787/575649abb-en>

Roberts et al. (2015). Biosafety research for non-target organism risk assessment of RNAi-based GE plants. *Frontiers in Plant Science*, 6, 958.
<https://doi.org/10.3389/fpls.2015.00958>

Bundesamt für
Naturschutz